# Thermal-physiological Strategies Underlying the Sympatric Occurrence of Three Desert Lizard Species

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**Abstract** Sympatric reptiles are the ideal system for investigating temperature-driven coexistence. Understanding thermally physiological responses of sympatric lizards is necessary to reveal the physiological mechanisms that underpin the sympatric occurrence of reptiles. In this study, we used three lizard species, *Eremias argus, E. multiocellata*, and *Phrynocephalus przewalskii*, which are sympatric in the Inner Mongolia desert steppe, as a study system. By comparing their resting metabolic rates (RMR) and locomotion at different body temperatures, we aimed to better understand their physiological responses to thermal environments, which may explain the sympatric occurrence of these lizards. Our results showed that *E. argus* had significantly higher RMR and sprint speed than *E. multiocellata*, and higher RMR than *P. przewalskii*. In addition, the optimal temperature that maximized metabolic rates and locomotion for *E. argus* and *E. multiocellata* was 36°C, whereas for *P. przewalskii* it was 39°C. Our study revealed the physiological responses to temperatures. *Eremias argus* and *E. multiocellata*, which have lower body temperatures than *P. przewalskii*, depend on higher RMR and locomotion to compensate for their lower body temperatures in field conditions. Our study also highlights the importance of using an integrative approach, combining behavior and physiology, to explore the basis of sympatric occurrence in ectothermic species.

Keywords Sympatric lizards, resting metabolic rate, locomotion, *Eremias argus, E. multiocellata, Phrynocephalus przewalskii* 

## 1. Introduction

Understanding the mechanisms that allow species coexistence is one of the core issues in community ecology. Niche differentiation is considered to be the basis of coexistence among sympatric species. Niche differentiation is a process by which competing species utilize environmental resources differently, and it can include aspects such as activity period, use of space, and food preferences (Hardin, 1960; Shurin *et al.*, 2004). Pacala and Roughgarden (1985), for example, reported the existence of several anole lizards that shared food resources (i.e., insects) on the Caribbean islands but occupied different microhabitats such as leaf litter floor and branches to avoid any possible competition for microhabitats or food resources. *Lophophorus sclateri* and *Ithaginis cruentus*, two pheasant species, were observed to share high-altitude habitats in Gaoligong Mountain but to feed on different plants or different parts of the same plants (Luo *et al.*, 2016).

In ectotherms, performances of specific activities such as locomotion, immunity, growth, and even reproduction are greatly affected by body temperatures (Angilletta, 2009; Angilletta *et al.*, 2002). Within thermal tolerance

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range, the physiological performance of ectotherms is known to enhance as body temperature increases until the optimal temperature is reached; after that, performance rapidly decreases if body temperature continues to increase (Huey and Kingsolver, 1989). Because of its effect on body temperature and thus on function and performance, environmental temperature is one of the most important ecological factors for ectotherm animals (Angilletta, 2009) and, therefore, is also considered an important resource (e.g., Li et al., 2017). Among sympatric ectotherms, different temperature preferences may result in different body temperatures, which in turn may result in segregation of microhabitat utilization and allow sympatric occurrence (Adolph, 1990; Hertz, 1992; Martinvallejo et al., 1995; Wilkinson and Grover, 1996). Understanding how variation in thermal environment affects sympatric species has become a hot topic in animal ecology (e.g., Li et al., 2017; Osojnik et al., 2013; Rusch et al., 2018; Žagar et al., 2015).

As ectotherms, sympatric lizards constitute ideal systems for investigating temperature-driven niche differentiation and coexistence (Pianka, 1986). Lizards can regulate their body temperatures in a narrow range mainly through behaviors to facilitate physiological functions (Adolph, 1990; Angilletta, 2009). On the Mongolian plateau, three ground-dwelling sympatric lizard species coexist in arid, semi-arid, or grass lands area: Eremias argus, E. multiocellata, and Phrynocephalus przewalskii (Zhao et al., 1999). These sympatric species have been demonstrated to occupy different microhabitats (e.g., vegetation coverage) and to have significantly different thermal preferences and active body temperatures. Phrynocephalus przewalskii mainly selects open and warm microhabitats and has a higher preferred body temperature range (T\_{sel}: 33.9–39.2  $^\circ\!\mathrm{C}$  ) than *E. argus* ( $T_{sel}$ : 32.8–37.5°C) and *E. multiocellata* ( $T_{sel}$ : 33.4–36.8°C), which occupy shaded and cool habitats (Table S1) (Li et al., 2017). Accordingly, the active body temperatures are significantly higher in *P. przewalskii* than in E. argus and E. multiocellata (Table S1) (Li et al., 2017). These species' demands for thermal environmental resources were speculated to be met because of microhabitat differentiation, which may be an important basis for their sympatric occurrence (Li et al., 2017). Nevertheless, as ectotherms, their body temperatures should be effective on fitness-related physiological functions before they affect their sympatric occurrences (Angilletta, 2009; Angilletta et al., 2002; Hochachka and Somero, 2002). Our current knowledge on these species' differences regarding thermal preference or active body

temperatures is insufficient to explain the effect of their thermal niche-partitioning on promoting their sympatric occurrence. Therefore, in order to reveal the physiological basis that underpins their temperature-driven sympatric occurrence, it is critical to investigate the effects of body temperature on important physiological traits of *E. argus*, *E. multiocellata*, and *P. przewalskii*.

Metabolism is one of the most important physiological processes because it determines an organism's demands from the environment and energy allocation among functions (Brown et al., 2004; McNab, 2002). Lizards' metabolic rates have been demonstrated to be significantly sensitive to body temperatures; these, in turn, require the animals' fast response to any environment thermal variation (e.g., Ma et al., 2018a; Ma et al., 2018b; Sun et al., 2018). For example, the metabolic rates were enhanced as body temperature increasing, and could respond to thermal variation by acclimation in lizard (e.g., Sun et al., 2018). Furthermore, the segregation of metabolic rates at the same temperatures may help niche differentiation (Žagar et al., 2018; Žagar et al., 2015). In addition, locomotion is thermal dependent and critical to escape danger, forage, and choose mating partners, and thus affects survival and reproduction in ectotherms (Robson and Miles, 2010; Shine, 2003; Shu et al., 2010; Wilson, 2001).

Here, with sympatric E. argus, E. multiocellata and P. przewalskii from Shierliancheng of Inner Mongolia area as a study system, we measured the metabolic rates and locomotion of each species at a range of body temperatures from 15-42°C in order to determine interspecific differences in physiology and fitness-related performances. We predicted that: 1) E. argus and E. multiocellata, which occupied shaded microhabitats and have lower active body temperatures, would have higher metabolic rates and sprint speeds but lower optimal temperatures for sprint speed than P. przewalskii at a same temperature within a moderate range before the optimal temperature is reached; 2) combined with temperaturerelated curves of metabolic rates and sprint speed, the three sympatric species would perform better when at their own active body temperatures under preferred microhabitat conditions.

#### 2. Materials and Methods

**2.1. Lizard collection and husbandry** The grounddwelling lizards *E. argus, E. multiocellata*, and *P. przewalskii* were collected from late June to early July at the Shierliancheng field station, Institute of Grassland

Research of the Chinese Academy of Agricultural Sciences (111°09' E, 40°21' N, elevation 1010-1021 m), where the ground-dwelling lizards (i.e., E. argus, E. multiocellata, and P. przewalskii) sympatry (Li et al., 2017). During collection, gravid female lizards were determined by palpate and released. Male and nongravid female adults of each species were transferred to our laboratory in Beijing, where they were weighed (to the nearest 0.001 g) and measured (to the nearest 1 mm). Every five or six lizards of each species were kept in a terrarium (550  $\times$  400  $\times$  350 mm<sup>3</sup>, length  $\times$  width  $\times$ height). The terraria were placed in a temperaturecontrolled room at 20°C with photoperiod cycles of 10 h dark: 14 h light (6:00–20:00). A heating bulb (50 w) was placed above one end of each terrarium to provide a temperature gradient inside the terrarium from 20-50°C within each photoperiod. Food (crickets dusted with vitamins) and water were provided ad libitum.

2.2. Resting metabolic rate After seven days of husbandry, the resting metabolic rates (RMR) of 14 E. argus (7  $\stackrel{\wedge}{\rightarrow}$  and 7  $\stackrel{\circ}{\rightarrow}$ ), 13 *E. multiocellata* (5  $\stackrel{\wedge}{\rightarrow}$  and 8  $\stackrel{\circ}{\rightarrow}$ ), and 14 *P. przewalskii* (7  $\stackrel{\frown}{\circ}$  and 7  $\stackrel{\bigcirc}{\circ}$ ) (total n = 41) were measured at ten test temperatures (15, 18, 21, 24, 27, 30, 33, 36, 39, and  $42^{\circ}$ C), with one temperature every other day in a random sequence. Lizards were fasted for at least 12 hours before each measurement. Initially, each lizard was placed in an incubator (Sanyo, Japan) at the selected test temperature for 90 min; then, the lizard was placed in a respirometry chamber within the incubator and the measurement was performed. RMR was measured using a closed-flow respirometry system with a volume of 281.4 ml (Stable Systems International Inc. Las Vegas, NV, USA) and estimated via CO<sub>2</sub> production rate following a previously established method (Sun et al., 2018). In brief, the system contained the following modules: Universal interface II (UI2), Subsampler TR-S (SS3), Mass flow control unit (MFC-2), Oxygen analyzer (FC-10a), and Carbon dioxide analyzer (Ca-10a). At the beginning of each measurement, the system was opened for two to three minutes so that air came through an entrance tube with a flow rate of 300 ml/min to make the baseline stable. Afterward, the measurement system was transferred to a closed-circuit respirometry, and then carbon dioxide production rates  $(V_{CO2})$  in closedcircuit were continuously recorded for 10 min. The entire environment was dark (no light exposure) during measurements, and all measurements were conducted from 10:00-18:00 to minimize the effects of circadian rhythms. The metabolic rates were calculated as the CO<sub>2</sub> production per gram<sup>0.75</sup> of body mass per hour (ml/g<sup>0.75</sup>/

hr) following the 'metabolic theory of ecology' (Brown *et al.*, 2004), with the equation of metabolic rates =  $V_{CO2} \times$  volume/body mass, where  $V_{CO2}$  is the CO<sub>2</sub> production rate in percentage (%/hr) in the closed circuit with the volume of 281.4 ml (Sun *et al.*, 2018). After each measurement, lizards were transferred back to the terraria until the next measurement initiated. After around 20 days of RMR tests, the locomotion tests were performed.

**2.3. Locomotion** The locomotion of 37 lizards was determined using a 1 000 mm custom-made race track at ten test temperatures randomly (15, 18, 21, 24, 27, 30, 33, 36, 39 and 42°C) after the RMR measurements. Locomotion measurements were not performed in four E. argus as they were out of conditions (i.e., inactive or unwilling to move under stimulation) one day after the RMR experiments. Before the locomotion measurement, lizards were placed in an incubator at each test temperature for 90 min. The sprint speed was then determined in a 1 000 mm race track with photoelectric timers every 200 mm. For measurements, the lizard was placed in one end of the track and then stimulated on the tail with a paintbrush to run along the race track. The time spent by the lizard to run over every 200 mm interval was recorded using the photoelectric timer. Tests were conducted from 10:00-14:00. Each individual was tested twice with an interval of one hour between tests; the fastest speed of the two tests (i.e., 10 speed records) was recorded as the sprint speed for each lizard. Tests were conducted every other day.

**2.4. Statistical analysis** Before statistical analysis, Kolmogorov-Smirnov test and *Levene's* test were conducted to detect data normality and variance homogeneity. Repeated-measures ANOVAs were conducted to determine species differences in RMR and locomotion, with species as a main factor and test temperatures as a repeated factor. When interaction between species and test temperatures was detected, a further comparison was conducted to analyze the differences among species at relevant test temperature ranges.

#### 3. Results

**3.1 Resting metabolic rates** The RMRs were significantly different among the three species. *Eremias argus* had a higher RMR than *E. multiocellata* and *P. przewalskii* (*E. argus*<sup>a</sup> > *E. multiocellata*<sup>b</sup> > *P. przewalskii*<sup>b</sup>;  $F_{2,38} = 3.65$ , P = 0.036) (Figure 1). RMR increased with test temperature until the optimal temperature was

reached; then, RMR decreased as temperature continued to increase (repeated factor 'test temperature':  $F_{9,342}$  = 97.956, P < 0.0001). The optimal temperature of RMR for *E. argus* and *E. multiocellata* was 36°C, whereas for *P. przewalskii* it was 39°C. The effect of the test temperatures on RMR was species-dependent. At low temperatures (from 15–36°C), *E. argus* had a significantly higher RMR than *P. przewalskii*, with *E. multiocellata* in between (*E. argus*<sup>a</sup> > *E. multiocellata*<sup>ab</sup> > *P. przewalskii*<sup>b</sup>;  $F_{2,38} = 8.251$ , P = 0.001). At high temperatures (39–42°C), the RMR of *P. przewalskii* was significantly higher than those of *E. argus* and *E. multiocellata* (*P. przewalskii*<sup>a</sup> >*E. argus*<sup>b</sup> > *E. multiocellata*<sup>b</sup>;  $F_{2,38} = 18.023$ , P < 0.0001) (Figure 1).

3.2. Locomotion Sprint speed of E. argus was significantly higher than that of *E. multiocellata*, whereas sprint speed of P. przewalskii was not significantly different from those of either species (*E. argus*<sup>a</sup> > *P. przewalskii*<sup>ab</sup> > *E. multiocellata*<sup>b</sup>;  $F_{2.34} = 5.376$ , P = 0.009) (Figure 2). Test temperature had a significant effect on sprint speed ( $F_{9,306} = 61.687$ , P < 0.0001). Sprint speed increased with temperature until the optimal temperature, after which it decreased as temperature continued to increase. The optimal temperature of sprint speed for E. argus and E. multiocellata was 36°C, whereas for P. przewalskii it was 39°C. At low temperatures (15–36°C), the sprint speed of *E. argus* was significantly higher than that of E. multiocellata; the sprint speed of P. przewalskii was not significantly different from those of either species (E.  $argus^{a} > P. przewalskii^{ab} > E. multiocellata^{b}; F_{2.34}$ = 4.439, P = 0.019). At high temperatures (39–42°C), the sprint speeds of P. przewalskii and E. argus were similar and both higher than that of E. multiocellata (P. przewalskii<sup>a</sup> > E. argus<sup>a</sup> > E. multiocellata<sup>b</sup>;  $F_{2,34} = 6.792$ , P = 0.003) (Figure 2).

### 4. Discussion

On the basis of previously known thermal biology traits such as active body temperatures, and thermal preference of the sympatric lizards *E. argus*, *E. multiocellata*, and *P. przewalskii* (Tables S1) (Li *et al.*, 2017), in the present study we determined the interspecific differences in RMR and locomotion at different body temperatures from 15 to 42°C. We found that *E. argus* have significantly higher RMR and locomotor performance, and *E. multiocellata* has significant higher RMR, when compared to *P. przewalskii*, especially at low body temperatures from 15 to 36°C, but lower optimal temperatures for RMR and locomotion, which indicates the existence



**Figure 1** Resting metabolic rate (RMR) of *Eremias argus, E. multiocellata*, and *Phrynocephalus przewalskii* at 15, 18, 21, 24, 27, 30, 33, 36, 39, and 42°C. RMR was expressed as CO<sub>2</sub> production per g<sup>0.75</sup> body mass per hour (ml/g<sup>0.75</sup>/hr). Red circles, blue triangles, and green rectangles indicate the RMR of *E. argus, E. multiocellata*, and *P. przewalskii*, respectively. The optimal temperatures for *E. argus, E. multiocellata*, and *P. przewalskii* were 36°C, 36°C, and 39°C, respectively. Data are shown as mean  $\pm$  SE. The RMRs (expressed as ml/g/hr) are also provided in Table S2, for the convenience of interspecific comparison in 'Meta-Analysis'.



**Figure 2** Locomotion of *Eremias argus*, *E. multiocellata*, and *Phrynocephalus przewalskii* at 15, 18, 21, 24, 27, 30, 33, 36, 39, and 42°C. Locomotion was expressed as sprint speed (m/s). Red circles, blue triangles, and green rectangles indicate the sprint speed of *E. argus*, *E. multiocellata*, and *P. przewalskii*, respectively. The optimal temperatures for *E. argus*, *E. multiocellata*, and *P. przewalskii*, are 36°C, 36°C, and 39°C, respectively. Data are shown as mean  $\pm$  SE.

of a physiological adaptation to the decrease in body temperature that also influences their performances. Therefore, we found that physiology and performance are both fine-tuned to the thermal preferences of the studied lizard species.

The interspecific differences in metabolic rates among *E. argus, E. multiocellata*, and *P. przewalskii* are consistent with the differences that were observed in hatchlings incubated under fluctuant temperatures (Ma et al., 2018a, 2018b). The metabolic rates of hatchlings of E. argus and E. multiocellata were higher than in those of P. przewalskii and enhanced with the increase in the test temperatures; this may indicate that the interspecific differences are fixed at the different life cycle stages of these species. As the fundamental physiological process, metabolic rates may reflect the organism's physiological response to the environment (Brown et al., 2004; McNab, 2002). However, unlike endotherms-which have thermal neutral zones within which organismal metabolic rates are thermally insensitive—, ectothermic metabolic rates are significantly thermally dependent, with a slow increasing rate at low temperatures and a rapid increase at high temperatures, followed by a steep drop after optimal temperature is reached (Gillooly et al., 2001; White, 2011). As the fundamental 'pacemaker' of biological rate, metabolic rate is related to lizard species' body temperature, which is in turn affected by thermal environments (Brown et al., 2004; Glazier, 2015). Eremias argus and E. multiocellata select shaded microhabitats and thus have lower body temperatures than P. przewalskii (Table S1) (Li et al., 2017). Having high metabolic rates at low temperatures (15-36°C, Figure 1) may create advantages for *E. argus* and *E.* multiocellata at low temperatures by allowing the allocation of more metabolic energy for the performance of activities such as escaping or foraging (e.g., Sun et al., 2018; White et al., 2012; White and Kearney, 2013). In contrast, as a higher RMR may enable a higher metabolic energetic production, a higher RMR at high temperatures in P. przewalskii may be responsible for improving this species' performance, accompanied by higher thermal preferences and active body temperatures. Alternatively, a higher RMR at high temperatures may induce more energetic allocation for maintenance, which might be a cost (Sokolova et al., 2012).

The existence of interspecific differences in locomotion across body temperatures has been demonstrated in numerous species of lizards (Chen *et al.*, 2003; Du *et al.*, 2000; Ji, 1995; Ji *et al.*, 1996; Sun *et al.*, 2014; Xu, 2001; Zhang and Ji, 2004). Locomotion could reflect the species' ability to escape, forage, and even reproduce (Husak and Fox, 2006; Shu *et al.*, 2010). The higher sprint speed in *E. argus* than in *P. przewalskii* and *E. multiocellata* may result in selective advantages in escaping, foraging, and even reproduction, especially at low temperatures (Bergmann and Irschick, 2010; Xu, 2001; Zamora-Camacho *et al.*, 2014).

As they are desert species, the optimal temperatures for sprint speed in E. argus (36°C), E. multiocellata (36°C), and P. przewalskii (39°C) are higher than those reported for skinks (32-34°C) (Du et al., 2000; Ji, 1995; Xu, 2001) and grass lizards (28-34°C) (Chen et al., 2003; Ji et al., 1996; Zhang and Ji, 2004). In addition, the high optimal temperature for P. przewalskii found in this study is consistent with this species' higher active body temperatures, thermal preference, and thermal tolerance (Li et al., 2017) if compared to the sympatric E. argus and E. multiocellata. The differences in optimal temperatures may be an evolutionary consequence of adaptation to different climatic environments (Zhang and Ji, 2004). Lizards in open and warm environments (e.g., deserts) tend to have high body temperatures, and their optimal temperatures for functions are normally positively correlated with body temperatures (e.g., Ji, 1995; Zhang and Ji, 2004). Potentially, active body temperatures and optimal temperatures for function may be both affected by habitat thermal environments through natural selection or acclimation (Gilbert and Miles, 2017; Sinclair et al., 2016). Within a species' distribution area, other climatic factors (i.e., humidity) may also directly or indirectly induce variation in thermal traits in lizards, including optimal temperatures of thermal performance curve or critical temperatures (Sinclair et al., 2016; Sunday et al., 2012). Nonetheless, in the present study, these three sympatric species lived in adjacent microhabitats with very similar precipitation and evaporation indices (Wang et al., 2016); thus, we propose that the divergences in optimal temperatures of metabolic rates and locomotion among species were driven by thermal environments. However, future studies on the effect of multiple factors on thermal traits in sympatric ectotherms systems would be very important and necessary.

In general, the sympatric species *E. argus*, *E. multiocellata*, and *P. przewalskii* have different preferred microhabitats, resulting in interspecies variation of thermal preferences and active body temperatures (Table S1) (Li *et al.*, 2017). *Eremias argus* and *E. multiocellata* resort to high physiological processes within a low temperatures range (i.e.,  $15-36^{\circ}$ C) to compensate for their lower body temperatures in field conditions. Alternatively, *P. przewalskii* occupies open and warm microhabitats, and it has higher body temperatures. Lower metabolic rates and sprint speed at low body temperatures may be an adaptive strategy for *P. przewalskii*; its low energetic production at low body temperatures results in a lower energetic cost—indicated by low sprint speed—, which in turn reduces energy expenditure (Brown *et al.*,

2004; Young et al., 2011). Similarly, within a certain geographical scale, populations or species that occupy warm habitats (e.g., tropical region) also have higher body temperatures, and therefore tend to have lower performance (e.g., swimming speed) and metabolism rates when compared to species from cold environments at even temperatures, as 'Metabolic Cold Adaptation' predicts (e.g., White et al., 2012). Given that, the combination of different active body temperatures and physiological trait responses is an effective solution that allows the sympatric occurrence of the three desert lizards, by enabling their functions within an active body temperature range. Our study highlights the importance of integrative investigations on temperature-driven sympatric occurrence at a physiological level, based on thermal biological traits and active body temperatures. Future studies should focus on the dynamic of the effects of body temperatures on physiological responses in field to reveal the modification of physiological and body temperatures to thermal variation. Especially in the context of climate warming, studies integrating body temperatures alterations and thermal performance curves may provide insight into the responses and the evaluation of the vulnerabilities of sympatric species (Sinclair et al., 2016).

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# Appendix

**Table S1** Average active body temperatures  $(T_a)$ , active body temperatures at open microhabitat  $(T_a \text{ open})$ , active body temperatures at shaded microhabitat  $(T_a \text{ shade})$ , and thermal preference  $(T_{sel})$  of *Phrynocephalus przewalskii*, *Eremias argus*, and *E. multiocellata*. Lizards were collected at the Shierliancheng field station, Institute of Grassland Research of the Chinese Academy of Agricultural Sciences (111°09′ E, 40°21′ N, elevation 1 010–1 021 m). Data are expressed as mean ± SE and follow Li *et al.* (2017).

|                        | Ν   | P. przewalskii | Ν  | E. argus         | Ν   | E. multiocellata |
|------------------------|-----|----------------|----|------------------|-----|------------------|
| $T_{\rm a}$ (°C)       | 129 | $38.1 \pm 0.2$ | 84 | 37.5 ± 0.2       | 116 | 36.6 ± 0.2       |
| $T_{\rm a}$ open (°C)  | 43  | $40.19\pm0.88$ | 29 | $39.07 \pm 0.67$ | 39  | 38.33 ± 0.14     |
| $T_{\rm a}$ shade (°C) | 40  | 35.83 ± 1.78   | 27 | 35.70 ± 1.69     | 32  | 34.81 ± 0.19     |
| $T_{(\circ C)}$        | 17  | $36.6\pm0.2$   | 21 | $35.3 \pm 0.2$   | 21  | $35.2 \pm 0.2$   |
| $T_{\rm sel}$ (°C)     | 17  | 33.9–39.2      | 21 | 32.8-37.5        | 21  | 33.4–36.8        |

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|                               | N  | RMR at 15°C                            | RMR at 18°C       | RMR at 21 °C      | RMR at 21 °C RMR at 24°C  | RMR at 27°C       | RMR at 30°C       | RMR at 33 °C                        | RMR at 36°C       | RMR at 39°C                         | RMR at 42°C       |
|-------------------------------|----|--|-------------------|-------------------|---|-------------------|-------------------|-------------------------------------|-------------------|-------------------------------------|-------------------|
| Eremias<br>argus              | 14 | $14 	0.045 \pm 0.003 	0.145 \pm 0.013$ | $0.145 \pm 0.013$ | $0.185 \pm 0.012$ | $0.185 \pm 0.012$ $0.368 \pm 0.033$ $0.436 \pm 0.042$ $0.579 \pm 0.048$ $0.642 \pm 0.057$ $0.909 \pm 0.066$ $0.687 \pm 0.031$ $0.407 \pm 0.031$ | $0.436 \pm 0.042$ | $0.579 \pm 0.048$ | $0.642 \pm 0.057$                   | $0.909 \pm 0.066$ | $0.687 \pm 0.031$                   | $0.407 \pm 0.031$ |
| Eremias multiocellata         | 13 | $0.077 \pm 0.015$                      | $0.150 \pm 0.017$ | $0.235 \pm 0.016$ | $0.282 \pm 0.024$   | $0.371 \pm 0.044$ | $0.445 \pm 0.044$ | $0.521 \pm 0.052$                   | $0.605 \pm 0.071$ | $0.545 \pm 0.067$                   | $0.224 \pm 0.011$ |
| Phrynocephalus<br>przewalskii | 14 | $0.109 \pm 0.011$                      | $0.121 \pm 0.007$ | $0.130 \pm 0.016$ | $0.130 \pm 0.016$ $0.182 \pm 0.021$   | $0.335 \pm 0.046$ | $0.428 \pm 0.053$ | $0.428 \pm 0.053$ $0.466 \pm 0.043$ | $0.650 \pm 0.060$ | $0.835 \pm 0.060$ $0.647 \pm 0.073$ | $0.647 \pm 0.073$ |

Table S2 Results of resting metabolic rate (RMR) of each species at different temperatures. RMR was calculated as the CO<sub>2</sub> production per gram of body mass per hour (ml/g/hr)<sup>\*</sup>. Data are shown

\* The RMR values are the same as those shown in the main text, but they are herein differently expressed.